

Role of Hypoxia in the Hallmarks of Human Cancer

Kai Ruan,¹ Gang Song,^{1,2} and Gaoliang Ouyang^{1*}

¹Key Laboratory of the Ministry of Education for Cell Biology and Tumor Cell Engineering, School of Life Sciences, Xiamen University, Xiamen 361005, China

²Cancer Research Center, Medical College, Xiamen University, Xiamen 361005, China

ABSTRACT

Hypoxia has been recognized as one of the fundamentally important features of solid tumors and plays a critical role in various cellular and physiologic events, including cell proliferation, survival, angiogenesis, immunosurveillance, metabolism, as well as tumor invasion and metastasis. These responses to hypoxia are at least partially orchestrated by activation of the hypoxia-inducible factors (HIFs). HIF-1 is a key regulator of the response of mammalian cells to oxygen deprivation and plays critical roles in the adaptation of tumor cells to a hypoxic microenvironment. Hypoxia and overexpression of HIF-1 have been associated with radiation therapy and chemotherapy resistance, an increased risk of invasion and metastasis, and a poor clinical prognosis of solid tumors. The discovery of HIF-1 signaling has led to a rapidly increasing understanding of the complex mechanisms involved in tumor hypoxia and has helped greatly in screening novel anticancer agents. In this review, we will first introduce the cellular responses to hypoxia and HIF-1 signaling pathway in hypoxia, and then summarize the multifaceted role of hypoxia in the hallmarks of human cancers. *J. Cell. Biochem.* 107: 1053–1062, 2009. © 2009 Wiley-Liss, Inc.

KEY WORDS: HYPOXIA; HYPOXIA INDUCIBLE FACTOR; CANCER; MICROENVIRONMENT; METASTASIS; METABOLISM

Tumorogenesis in human is a multistep process that involves the sequential acquisition of a number of genetic, epigenetic, or somatic alterations as a result of increasing genomic instability caused by defects in cell cycle checkpoint controls [Hanahan and Weinberg, 2000]. These alternations enable cancer cells to acquire characteristics different from normal cells: resistance to growth inhibitory factors, proliferation in the absence of exogenous growth factors, evasion of apoptosis, limitless replication potential via the reactivation of telomerase, abnormal angiogenesis, evasion of destruction by the immune system, invasion and metastasis [Hanahan and Weinberg, 2000; Bao et al., 2004]. In addition to the genetic, epigenetic, or somatic changes that occur in cancer, the tumor microenvironment is now considered to be a critical factor in malignancy progression and metastasis, and it influences the response to conventional anti-tumor therapies [Hanahan and Weinberg, 2000; Roskelley and Bissell, 2002]. As one of the most pervasive microenvironmental stresses and common features of solid tumors, hypoxia plays an important but complex role in mediating or regulating each of these hallmarks in the progression of human tumors from microinvasive to metastatic cancers in vivo (Fig. 1). Herein, we focus on the role of hypoxia, especially the hypoxia-inducible factor-1 (HIF-1) pathway, on the hallmarks of human cancers.

HYPOXIA AND THE HIF-1 SIGNALING PATHWAY

THE CELLULAR RESPONSES TO HYPOXIA

Mammalian cells undertake a variety of responses to maintain oxygen homeostasis, a precise balance between the need for oxygen as an energy substrate for oxidative phosphorylation and other essential metabolic reactions and the inherent risk of oxidative damage to cellular macromolecules. Oxygen is only able to diffuse 100–180 μm from the end of the nearest capillary to cells before it is completely metabolized [Powis and Kirkpatrick, 2004]. However, rapidly growing tumors quickly outstrip the vascular supply and, thus, result in a poorly vascularized microenvironment characterized by hypoxia, low pH, and nutrient starvation [Pouyssegur et al., 2006; Denko, 2008]. Therefore, intratumoral hypoxia occurs when cells are located greater than this distance from a functional blood vessel for adequate diffusion of oxygen as a result of rapid tumor cell proliferation and abnormal blood vessels [Semenza, 2007a; Bertout et al., 2008]. To survive and grow in this hypoxic microenvironment, tumor cells co-opt adaptive mechanisms to switch to a glycolytic metabolism, promote proliferation, become resistant to apoptosis, obtain unlimited replication potential and genomic instability, evade immune attack, induce angiogenesis, and migrate to less hypoxic areas of the body (Fig. 1).

Grant sponsor: National Natural Science Foundation of China; Grant numbers: 30400239, 30570935, 30871242.

*Correspondence to: Dr. Gaoliang Ouyang, Key Laboratory of the Ministry of Education for Cell Biology and Tumor Cell Engineering, School of Life Sciences, Xiamen University, Xiamen 361005, China. E-mail: oygldz@yahoo.com.cn

Received 10 February 2009; Accepted 21 April 2009 • DOI 10.1002/jcb.22214 • 2009 © Wiley-Liss, Inc.

Published online 28 May 2009 in Wiley InterScience (www.interscience.wiley.com).

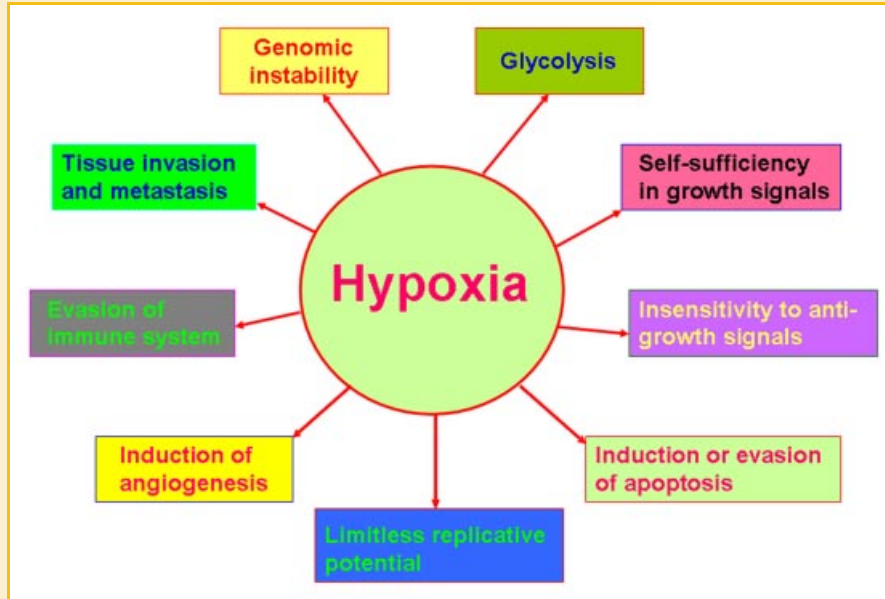


Fig. 1. The role of hypoxia in the hallmarks of human cancer. Under hypoxic microenvironments, cancer cells co-opt adaptive mechanisms to switch to a glycolytic metabolism, promote proliferation, induce or evade apoptosis, obtain unlimited replication potential and genomic instability, evade immune attack, induce angiogenesis, and invade and metastasize. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]

HIF-1 SIGNALING PATHWAY IN HYPOXIA

Hypoxia-inducible factors (HIFs) play an essential role in the maintenance of oxygen homeostasis in metazoan organisms and primarily mediate adaptive responses to reduced oxygen levels [Semenza, 2007b; Kaelin and Ratcliffe, 2008].

HIFs are composed of two subunits, HIF- α and HIF- β (also known as ARNT). Mammalian HIF- α subunits are encoded by three genes: *HIF-1 α* , *HIF-2 α* , and *HIF-3 α* . HIF-1 α is ubiquitously expressed, whereas HIF-2 α and HIF-3 α exhibit a more restricted tissue distribution. HIF-1 β is constitutively expressed and is largely insensitive to changes in oxygen levels, whereas the levels of all three HIF- α subunits are acutely regulated by hypoxia. HIF-1, a heterodimer of HIF-1 α and HIF-1 β , is overexpressed in various human cancers and has been recognized as one of the master regulators of oxygen homeostasis and mediates a wide variety of responses to hypoxia in tumorigenesis [Zhong et al., 1999; Bertout et al., 2008]. Current reports have shown that hypoxia activates HIFs by regulating two major switches that converge on α subunits. The first molecular switch regulates the overall cellular level of HIF- α while the second one controls transcriptional activities of HIF- α [for review see Kaluz et al., 2008]. There are two types of oxygen sensors, prolyl hydroxylase (PHD) proteins and factor inhibiting HIF-1 (FIH), to ensure full repression of the HIF pathway in well-oxygenated cells by regulating both the destruction and inactivation of HIF- α subunits [Jaakkola et al., 2001; Lando et al., 2002; Pouyssegur et al., 2006; Lisy and Peet, 2008]. In the presence of O₂, HIF- α is earmarked for degradation by hydroxylation which is catalyzed by PHDs, and then interact with the ubiquitin E3-containing ligase von Hippel-Lindau (pVHL). This interaction causes HIF- α to be ubiquitylated and then the polyubiquitylated HIF- α is targeted to the proteasome for degradation [Ohh et al., 2000; Kaelin and

Ratcliffe, 2008; Kaluz et al., 2008]. However, increases in the HIF- α isoform under hypoxia are largely due to the regulation of protein degradation. In the absence of oxygen, the oxygen sensor proteins PHDs and FIH are inactive because of the lack of available oxygen. HIF- α is stable and translocates to the nucleus and heterodimerizes with constitutively expressed HIF- β . The heterodimer then interacts with cofactors and binds to the hypoxia-response elements (HREs) in the promoters or enhancers of hypoxia-responsive genes which are crucial to mediate systemic hypoxia responses [Bertout et al., 2008; Chandel and Simon, 2008; Simon and Keith, 2008] (Fig. 2).

ROLE OF HYPOXIA IN THE HALLMARKS OF CANCER

RESISTANCE TO ANTI-PROLIFERATION SIGNALS AND INDEPENDENCE FROM EXOGENOUS GROWTH FACTOR SIGNALS

It is well established that cell proliferation is strictly regulated by the concerted action of both pro-proliferative and anti-proliferative signals. On the other hand, cells also show a remarkable regulatory flexibility that allows them to thrive under different external microenvironments. Cells launch various signaling pathways in response to external microenvironment alterations and in order to coordinate cell growth with stress responses [Lopez-Maury et al., 2008]. To continue growing under hypoxic conditions, cancer cells adapt to the absence of exogenous mitogenic growth signals and become resistant to the anti-proliferative signals [Harris, 2002].

In a hypoxic microenvironment, cancer cells can grow more independent of exogenous mitogenic growth signals than normal cells by overexpressing the mitogenic growth factors themselves or constitutively activating the downstream pathways of these growth factors. Hypoxia can induce cancer cells to express various growth

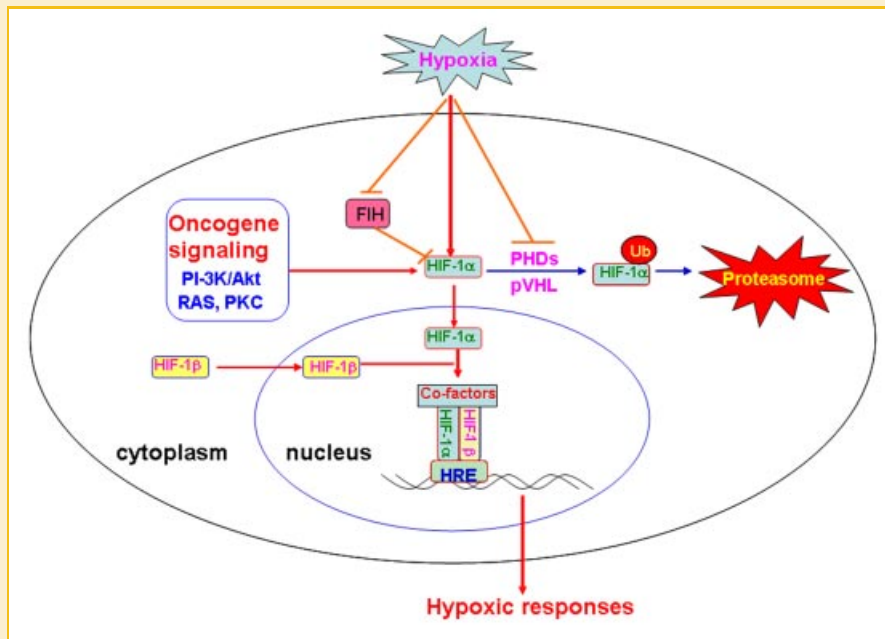


Fig. 2. Hypoxia signaling through HIF-1 pathways in human cancer. Two oxygen sensors, prolyl hydroxylase domain (PHD) proteins and factor inhibiting HIF-1 (FIH), ensure full inhibition of HIF-1 α in well-oxygenated cells by regulating the destruction and inactivation of HIF-1 α , respectively. Under hypoxic conditions, the oxygen sensor proteins PHDs and FIH are inactive because of the lack of available oxygen, and HIF-1 α protein hydroxylation and pVHL association are decreased. Hypoxic cells redirect HIF-1 α away from ubiquitylation and being targeting to the proteasome for degradation toward signaling to the nucleus. After translocation to the nucleus, HIF-1 α heterodimerizes with the constitutively expressed HIF-1 β subunit, and the heterodimer interacts with cofactors and binds to DNA at hypoxia-response elements (HRE) in the promoters or enhancers of numerous hypoxia-responsive genes. HIF-1-mediated transcriptional regulation is synergistically enhanced by cofactors such as p300/CBP, SRC-1, and Ref-1. Hypoxia-inducible genes regulate various biological processes, including cell proliferation, apoptosis, angiogenesis, metabolism, and metastasis. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]

factors such as EGF, insulin, IGF-1, IGF-2, and PDGF, which are known to promote cell proliferation [Maxwell et al., 2001; Powis and Kirkpatrick, 2004]. p42/p44 MAPK has been reported to phosphorylate HIF-1 α and activate the transcription of HIF-1-response genes to regulate cell proliferation [Richard et al., 1999]. Moreover, hypoxia-induced HIF-1 α activates transcription of vascular endothelial growth factor (VEGF) and VEGFR1 to promote endothelial cell proliferation and blood vessel formation [Harris, 2002].

In addition to taking advantage of various hypoxia-induced growth-promoting signaling, cancer cells also modify growth-inhibitory signaling events. For example, tumor suppressor PTEN can inhibit the PI-3K/Akt pathway and regulate cell growth and survival. PTEN is deleted or mutated in some human cancers. Restoration of wild-type PTEN in glioblastoma cell lines lacking functional PTEN ablates hypoxic induction of HIF-1-regulated genes. Thus, PTEN mutations may promote tumor growth by synergistically promoting HIFs-mediated responses [Zundel et al., 2000; Harris, 2002].

INDUCTION OR EVASION OF APOPTOSIS

Tumors grow in an uncontrolled manner through the imbalance of cell proliferation and death. The role of deregulation of apoptosis in tumorigenesis is complex and associated to successive and interdependent genetic and epigenetic events that gradually result in tumor formation [Nelson et al., 2004]. In contrast to normal cells, cancer cells can break the balance between pro- and anti-apoptotic

factors to promote cell survival under adverse environmental conditions, such as hypoxic stress, radiation, or chemotherapy [Semenza, 2002; Bao et al., 2004; Blagosklonny, 2004]. The mechanisms of apoptosis regulation under hypoxia are, however, not fully understood.

There is evidence that the mitochondrial permeability transition is a central mechanism in hypoxia-mediated apoptosis. HIF-1 α promotes the release of cytochrome c from mitochondria into the cytoplasm, and this p53-dependent apoptosis is mediated by Apaf-1 and caspase-9 [Soengas et al., 1999]. This process can be counteracted by members of the Bcl-2 family of anti-apoptotic molecules, such as Bcl-2 itself and Bcl-xL. Evidence has also been provided that Bax, another member of the Bcl-2 family, enhances hypoxia-induced apoptosis. HIF-1 activates BNIP3 and NIX, which induce a mitochondrial-pore permeability transition and cell death via a mechanism that does not involve cytochrome c release or caspases [Sowter et al., 2001; Harris, 2002]. In addition, the JNK pathway also contributes to hypoxia-induced apoptosis [Kunz et al., 2001]. Furthermore, only the phosphorylated HIF-1 α binds HIF-1 β under hypoxia, while the dephosphorylated form of HIF-1 α binds p53 and exerts pro-apoptotic effects [Suzuki et al., 2001]. Therefore, the phosphorylation status of HIF-1 α under hypoxia is a critical factor for the decision on whether HIF-1 promotes apoptosis. In addition, the effect of HIF-1 α on apoptosis is dependent on the cancer type and the presence or absence of genetic alterations that alter the balance between pro- and anti-apoptotic factors [Semenza, 2003].

However, hypoxic exposure can also increase resistance to apoptosis. Due to the lack of effective and adequate blood vessels in the early stages of tumor development, fast growing tumors often become hypoxic. Although hypoxia is toxic to both cancer cells and normal cells, cancer cells can undertake genetic and adaptive changes in response to hypoxia that permit them to be refractory to apoptosis and increase their ability to survive and proliferate compared with normal cells [Harris, 2002; Powis and Kirkpatrick, 2004]. Tumor cells have developed various mechanisms for escaping HIF-1-mediated apoptosis under a hypoxic microenvironment [Dong et al., 2001]. Zhang et al. have reported that, in some rodent cells, hypoxia can up-regulate the expression of the p53-negative regulator MDM2 and increase cells resistance to apoptosis *in vivo* and promote metastasis [Zhang and Hill, 2004]. Hypoxia can induce the expression of the anti-apoptosis protein IAP-2 or apoptosis repressor with caspase recruitment domain (ARC) [Schmid et al., 2004]. β -Catenin can enhance HIF-1-mediated transcription, thereby promoting cell survival and adaptation to hypoxia in colorectal tumorigenesis [Kaidi et al., 2007]. Moreover, the PI-3K/Akt survival pathway, a critical regulator of cell survival and proliferation, has been shown to be activated in various cell types under hypoxia [Chen et al., 2001; Song et al., 2005; Furuta et al., 2008; Wai and Kuo, 2008; Zeng et al., 2008; Walsh et al., 2009]. Our previous work has revealed that periostin, a secreted protein, can dramatically enhance metastatic growth of colon cancer by both preventing hypoxia-induced apoptosis in cancer cells and augmenting endothelial cell survival via the Akt/PKB pathway [Bao et al., 2004]. Osteopontin (OPN) is another secreted protein that plays an important role in the progression of tumor development [Rangaswami et al., 2006; Wai and Kuo, 2008]. Our recent results suggested that the underlying mechanism of OPN-mediated promotion of tumor development is largely associated with the ability of Akt activation to enhance cell survival under stress [Song et al., 2008a,b]. In this regard, acquired expression of periostin, OPN, and similar types of matricellular proteins may enable tumor cells to thrive in the hypoxic microenvironment [Ouyang et al., 2009; Ruan et al., 2009]. In addition, hypoxia acts as a selective pressure during tumor growth, eliminating cells with wild-type p53 and selectively and clonally expanding cells with mutant or otherwise inactive p53; these results indicate that hypoxia-mediated selection for p53 mutant cells with diminished apoptotic potential in solid tumors may account for the high prevalence of p53 mutations in human cancers [Graeber et al., 1996]. Therefore, hypoxia may provide a physiological selective pressure in tumors for the preferential expansion of variants that have lost their apoptotic potential [Graeber et al., 1996; Bao et al., 2004].

LIMITLESS REPLICATIVE POTENTIAL

Human somatic cells normally have a finite replicative potential. After a limited number of cell divisions, cells undergo senescence. Cell senescence inhibits the development of cancer by suppressing the proliferation of damaged or stressed cells that are at risk for malignant transformation [Campisi, 2005a]. Senescence-inducing stimuli are potentially oncogenic; therefore, cancer cells have to acquire alterations that allow them to avoid telomere-dependent and oncogene-induced senescence [Hanahan and Weinberg, 2000;

Hahn and Weinberg, 2002; Campisi and d'Adda di Fagagna, 2007; Gillies and Gatenby, 2007]. Current reports have revealed that cancer cells can overcome this obstacle, fail to senesce, and then become immortalized [Artandi and DePinho, 2000; Campisi, 2005b; Yaswen and Campisi, 2007]. Immortalization is an essential prerequisite for the formation of a cancer cell. Human cancer cells acquire the property of immortalization through maintenance of telomeres by either up-regulating the telomerase or alternatively lengthening the telomeres [Hanahan and Weinberg, 2000; Hahn and Weinberg, 2002].

Human telomerase (hTERT) has been identified as one of the key effectors in cell senescence. Under a hypoxic environment, telomerase activity in cancer cells increases and promotes cellular immortalization, and the increased telomerase activity is mediated by MAPK activation [Seimiya et al., 1999]. Negative regulation of HIF-1 by PDH2, FIH, HIF-1 siRNA, and the HIF-1 inhibitor YC-1 can elicit massive endometrial cancer cell senescence. hTERT expression is decreased in senescence that is induced by PDH2-mediated HIF-1 α down-regulation [Kato et al., 2006]. A recent study has shown that HIF-1 α -dependent macrophage migration inhibitory factor (MIF) expression is necessary for hypoxia-induced evasion from cell senescence, and MIF is required for HIF-1 α stabilization induced by hypoxia and PHD inhibitors [Winner et al., 2007]. Mouse embryo fibroblasts (MEFs) harboring pVHL inactivation show increased rates of growth arrest and the accumulation of markers associated with senescence [Young et al., 2008]. However, neither hypoxia induction nor treatment with PHD inhibitors is sufficient to induce senescence in wild-type of MEFs [Kim and Sharpless, 2008; Young et al., 2008]. Therefore, the senescence induced by pVHL loss is independent of its ability to downregulate HIF.

INDUCTION OF ANGIOGENESIS

It is well established that local growth and metastasis of a large variety of malignant tumors are dependent on the formation of new blood vessels. To grow beyond 1 to 2 mm in diameter, solid tumors require a blood supply to provide enough nutrients and oxygen. Interestingly, tumor development is associated with both increased microvascular density and an intratumoral hypoxic microenvironment. These paradoxical characteristics arise because the tumor vasculature is structurally and functionally abnormal, resulting in abnormal perfusion that is characterized by marked spatial and temporal heterogeneity.

Tissue hypoxia can induce a number of angiogenic factors that promote angiogenesis, such as VEGF, IL-8, angiogenin, FGF, and PDGF. VEGF plays a major role in physiological blood vessel formation and pathological angiogenesis in tumor growth [Ellis and Hicklin, 2008]. Up-regulation of VEGF and other factors such as angiopoietin 2 (Ang-2) promotes the initiation and progression of angiogenesis [Holash et al., 1999; Dewhirst et al., 2008]. VEGF-A is expressed in most cells, and attracts and guides sprouting neovessels into oxygen-depleted areas in tumor tissues [Pouyssegur et al., 2006]. Hypoxia can promote HIF-1 activity to up-regulate VEGF as a trigger for the initiation of angiogenesis but can also decrease the activity of angiogenic inhibitor thrombospondin to create a pro-angiogenic microenvironment [Laderoute et al., 2000; Dewhirst et al., 2008]. Our previous data [Bao et al., 2004] have shown that the

secreted protein periostin can dramatically enhance the ability of human microvessel endothelial cells to survive under hypoxic conditions, which is compatible with the survival-promoting effect induced by the potent angiogenic factor VEGF. Ang-2 is another key angiogenic factor induced by hypoxia, however, the precise mechanism of regulation of Ang-2 expression in hypoxia remains to be defined [Maisonpierre et al., 1997; Pouyssegur et al., 2006].

Based on currently available evidence, there are two different models, the hypoxic crisis model and the acceleration model, that explain the role of hypoxia during initiation or acceleration of angiogenesis [Holash et al., 1999; Cao et al., 2005; Dewhirst et al., 2008]. In the vascular crisis model, hypoxia can induce VEGF to promote the initiation of angiogenesis. However, the acceleration model proposes that hypoxia is not responsible for initiation of angiogenesis and that this initiation is driven by non-hypoxia-mediated mechanisms, such as inducing VEGF by oncogene activation. Once angiogenesis is initiated, cancer cells grow quickly and result in hypoxia. HIF-1 is then increased to accelerate angiogenesis [Dewhirst et al., 2008]. Taken together, hypoxia may promote angiogenesis in tumors in two ways: first, by directly stimulating the expression of a series of angiogenic factors involved in the growth and survival of endothelial cells and second, by inducing a microenvironment favorable for endothelial cells and cancer cell survival, migration, and invasion.

EVASION OF THE IMMUNE SYSTEM

The tumor microenvironment is regulated not only by cancer cells but also by endothelial cells, immune cells, and stromal elements [Shi et al., 2004]. The co-existence of cancer cells and anti-tumor immune cells ("Hellstrom Paradox") has been a challenging paradox for a long time [Lukashev et al., 2007]. Interestingly, cancer cells undergo various changes and acquire the ability to evade the immunosurveillance. However, the complex mechanisms by which cancer cells evade the immune attack under hypoxic microenvironment are poorly understood. One possibility may be that cancer cells secrete various immunosuppressive factors by HIF-1 α -dependent or -independent pathways to protect cancer cells from immune damage.

In addition to cancer cells, immune cells are also often exposed to low oxygen tensions and must adapt to hypoxia to function in tumors and other microenvironments that are located far from the ends of capillaries [Sitkovsky and Lukashev, 2005; Rius et al., 2008]. Monocytes are continually recruited into tumors, differentiate into tumor-associated macrophages, and then accumulate in hypoxic areas. Hypoxia is a critical intratumoral signal that stimulates tumor-associated macrophages to secrete a series of mitogenic factors, proangiogenic cytokines, and immunosuppressive agents by inducing such transcription factors as HIF-1 and HIF-2 [Talks et al., 2000; Lewis and Murdoch, 2005]. Hypoxia also inhibits the ability of macrophages to phagocytose dead or dying cells, present antigens to T cells, and inhibit the anti-tumor effects of macrophages [Murata et al., 2002; Lewis and Murdoch, 2005]. In addition, HIF-1 α plays a critical role in the survival of T cells by preventing them from undergoing activation-induced cell death in a hypoxic microenvironment [Makino et al., 2003; Sitkovsky and Lukashev, 2005]. Tumor cells are protected from immune attack in hypoxic and

immunosuppressive tumor microenvironments due to the inhibition of anti-tumor T cells. The HIF-1 α -mediated anti-inflammatory pathway and hypoxia-induced extracellular adenosine may cooperatively contribute to the tumor-protecting immunosuppressive effects of tumor hypoxia [Sitkovsky and Lukashev, 2005; Lukashev et al., 2007].

These observations support the hypothesis that not only malignant cells but also immune cells adapt to hypoxic microenvironments and promote the expression factors that are necessary to escape immunosurveillance; together, these data suggest that eliminating these signaling from tumor cells enhances the host anti-tumor immune response.

TISSUE INVASION AND METASTASIS

Invasion and metastases are the most common causes of morbidity and mortality induced by cancer. Tumor invasion and metastasis consist of multiple and complicated processes, including cellular disengagement and motility from the local microenvironment, degradation of the surrounding extracellular matrix, and cellular movement, all of which must be successfully completed to permit the outgrowth of metastatic tumors in the new microenvironment [Chambers et al., 2002; Murata et al., 2002; Bao et al., 2004; Ma et al., 2008; Nguyen et al., 2009]. Successful adaptation to varying microenvironmental constraints plays a critical role during tumorigenesis [Joyce and Pollard, 2009]. In the hypoxic environment, tumor cells undertake a series of changes not only to survive and grow in hypoxic microenvironments but also to subsequently expand and promote invasion and metastasis. Clinical studies have shown that tumor hypoxia is one of the important microenvironmental determinants for tumor cell dissemination. Adaptation to hypoxia may represent one of the key events during the transition from in situ to invasive breast cancer [Gatenby et al., 2007]. Another recent work has shown that HIF-1 α is a critical regulator of metastasis in a transgenic model of cancer initiation and progression [Liao et al., 2007]. Hypoxic tumor cells are more aggressive, prone to distant metastases, and tolerant to anti-cancer therapy. One report revealed that antisense HIF-1 α treatment can inhibit the expression of survivin and β 1 integrin, enhance apoptosis in human pancreatic cancer cells, and restrain the invasion and metastasis of pancreatic cancer [Chang et al., 2006]. Hypoxia or HIF-1 α overexpression promotes matrigel invasion by HCT116 human colon carcinoma cells, whereas this process is inhibited by HIF-1 α siRNA [Krishnamachary et al., 2003]. However, the mechanisms that result in the increased metastatic potential of tumor cells exposed to hypoxia and the exact role of HIF-1 α in the metastasis still have not been well defined.

It was proposed that the initial steps in metastasis involve epithelial-mesenchymal transition (EMT) [Yang et al., 2004; Polyak and Weinberg, 2009]. The well-polarized, adhesive epithelial cells are converted to non-polarized mesenchymal cells. EMT is a multi-step process that requires the coordination of multiple cellular events, including the disruption of intercellular adhesion mediated by cadherins at adherens junction, the loss of apicobasal polarity, cytoskeletal architecture reorganization, and the degradation of the basement membrane. Both hypoxia and overexpression of HIF-1 α have been shown to promote EMT and metastatic phenotypes

[Krishnamachary et al., 2006; Peinado et al., 2007; Yang et al., 2008]. Hypoxia-induced HIFs activation is associated with a concomitant loss of E-cadherin, one of the landmarks of invasion and a crucial feature of EMT [Beavon, 1999]. Hypoxia may attenuate the expression of E-cadherin via activation of the lysyl oxidase (LOX)-Snail pathway to promote tumor invasion and metastasis, indicating that hypoxia-induced LOX and HIFs are important factors that regulate tumor microenvironments to favor metastasis [Imai et al., 2003; Erler et al., 2006; Pouyssegur et al., 2006; Croker and Allan, 2008]. Notch signaling and Wnt pathways are also required to convert the hypoxic stimulus into EMT, increased motility, and invasiveness [Jiang et al., 2007; Sahlgren et al., 2008]. All these data indicate that tumor hypoxia and/or HIF signaling are strongly associated with malignant progression.

GENOMIC INSTABILITY

It is well known that genomic instability is responsible for cellular changes that confer progressive transformation on cancer cells [Hanahan and Weinberg, 2000; Huang et al., 2007; Shen et al., 2008]. Genotoxic stress activates cell cycle checkpoints and delays cell cycle progression to allow for DNA repair [Bao et al., 2001]. Genetic defects in DNA repair mechanisms and cell cycle checkpoints result in increased genomic instability and cancer predisposition. Emerging evidence indicates that hypoxia as a tumor microenvironmental stress can drive genomic instability through increased chromosomal rearrangement, gene amplification, and induction of intrachromosomal fragile sites in hypoxic cells [Coquelle et al., 1998; Bristow and Hill, 2008]. The frequency of point mutations in tumorigenic cells cultured under severe hypoxic treatment is 3.4-fold higher than that observed in normally oxygenated cells [Hockel and Vaupel, 2001]. Current studies have shown that hypoxia can suppress the expression of DNA mismatch repair (MMR) genes and result in increased mutagenesis [Bindra et al., 2005, 2007; Huang et al., 2007]. The loss of MMR renders human colon carcinoma cells hypersensitive to the ability of hypoxia to induce microsatellite instability and generate highly drug-resistant clones in the surviving population [Kondo et al., 2001]. Hypoxia contributes further to genetic instability by HIF-1 α dependent transcriptional repression of the mismatch repair genes MSH2 and MSH6 [Koshiji et al., 2005]. In addition, polyploidy is also much more common in hypoxia. Hypoxia can decrease the expression levels of Brca1 and Rad51, which are two important mediators of homologous recombination in mammalian cells [Bindra et al., 2005, 2007]. Hypoxia also regulates cell cycle checkpoints and DNA repair in the prostate epithelium, thereby driving genomic instability and tumor aggression [Chan et al., 2007]. Furthermore, several DNA damage and DNA repair checkpoints proteins, including ataxia-telangiectasia mutated (ATM), ATM- and Rad3-related (ATR), Chk1, Chk2, Brca1, and p53, are activated in response to hypoxic exposure [Hammond et al., 2002, 2003; Gibson et al., 2005, 2006; Bindra et al., 2007].

GLYCOLYSIS IN HYPOXIC CELLS

In addition to the roles of hypoxia and the HIF-1 pathway in directly and indirectly mediating and regulating these above hallmarks of cancer, the adaptive shift from aerobic to anaerobic metabolism can

also be regarded as one of the important hallmarks of cancer cells under hypoxia [Gillies and Gatenby, 2007; Gatenby and Gillies, 2008]. The above essential hallmarks of cancer are intertwined with an altered cell-intrinsic metabolism, either as a consequence or as a cause [Kroemer and Pouyssegur, 2008].

Normal tissues and cells in well-oxygenated environments typically rely on highly efficient aerobic metabolism to generate ATP [Kim et al., 2007; Fang et al., 2008; Aragonés et al., 2009]. Under hypoxic conditions, however, cancer cells adjust their cellular physiology and metabolism to the new microenvironment and switch their glucose metabolism from the oxygen-dependent oxidative phosphorylation to the oxygen-independent glycolysis. There is abundant evidence that components of the glycolytic pathway are up-regulated in cancer cells. Hypoxic cancer cells use glycolysis as a primary mechanism of ATP production, and HIFs are master regulators of glucose metabolism during hypoxia [Harris, 2002; Semenza, 2003; Brahimi-Horn et al., 2007; Denko, 2008]. Recently, it was revealed that HIF-1 mediates this adaptation to hypoxia in cancer cells by down-regulating tricarboxylic acid (TCA) cycle activity and mitochondrial oxygen consumption through inhibition of pyruvate dehydrogenase [Kim et al., 2006; Papandreou et al., 2006]. HIF-1 can inhibit mitochondrial biogenesis and cellular respiration and increase glycolysis in VHL-deficient renal cell carcinoma by decreased repression of c-Myc activity [Zhang et al., 2007]. HIF-1 and c-Myc collaborate to activate hexokinase 2 (HK2) and pyruvate dehydrogenase kinase 1 (PDK1), resulting in increased conversion of glucose to lactic acid [Dang et al., 2008]. Furthermore, mammalian target of rapamycin (mTOR) signaling represents another critical regulator of cellular metabolism and is regarded as a positive regulator of glycolysis as well as of the biosynthesis of amino acids, proteins, lipids, and nucleotides [Wouters and Koritzinsky, 2008]. Therefore, HIFs may collaborate with Myc and mTOR in both synergistic and antagonistic ways to regulate cellular metabolism downstream of some of the most common mutations in cancer [Zhang et al., 2007; DeBerardinis et al., 2008; Wouters and Koritzinsky, 2008]. However, the mystery behind the molecular mechanisms of this metabolic switch under hypoxia is still not well understood.

CONCLUDING REMARKS

Tumorigenesis largely depends on alterations in the heterotypic interactions between incipient cancer cells and their normal neighbors. Virtually all types of human tumors are composed of several distinct cell types, such as cancer cells, immune cells, fibroblasts, and endothelial cells [Hanahan and Weinberg, 2000]. The evolution of a clinically significant invasive cancer from a focus of microinvasion requires not simply tumor mass growth but rather an active collaboration of malignant epithelial cells and normal mesenchymal cells [Gatenby and Gillies, 2008]. Therefore, it is widely accepted that tumors can be regarded as complex tissues in which cancer cells have conscripted and subverted normal cells to serve as active collaborators in their neoplastic process [Hanahan and Weinberg, 2000; Gatenby and Gillies, 2008]. To survive and grow in these complex microenvironments, cancer cells have to

overcome many noxious stimuli, such as hypoxia, that may induce cell death. However, it is not well understood how cancer cells adapt and manipulate pathways to cooperate with other cell types in tumors to promote their own survival and growth under hypoxic microenvironments. Current reports have revealed the microRNA signature of hypoxia in cancer cells, but the mechanisms by which hypoxia regulates the microRNA levels and the role of microRNAs in eliciting hypoxia responses are not well known [Kulshreshtha et al., 2007; Camps et al., 2008; Ivan et al., 2008]. Recently, it has been recognized that cancer stem cells also play critical roles in tumorigenesis and that hypoxia-induced transcription factors may drive tumor growth through the generation or expansion of cancer stem cells [Bao et al., 2006; Keith and Simon, 2007; Rich and Bao, 2007; Croker and Allan, 2008]. However, the role of hypoxia in cancer stem cells in tumorigenesis remains less clear. Likewise, little is known about the precise mechanisms that are responsible for the limitless replication potential and genomic instability characteristics of hypoxia on cancer cells. Another important area will be to establish the molecular basis for how cancer cells switch their metabolism to adapt to hypoxia.

As one of the most pervasive microenvironmental stresses that can impact malignant progression, tumor hypoxia can be found in almost every solid tumor and be now widely recognized as a cause of treatment failure and poor outcome for a wide variety of adult malignancies and, thus, needs to be taken into account when evaluating prognostics and therapeutic options for cancer patients [Lundgren et al., 2007; Le and Courter, 2008]. HIF-1 inhibition may represent a global strategy for targeting the hypoxic tumor microenvironment and there is an extensive effort involved in identifying new more potent and specific HIF-1 inhibitors. However, HIFs-independent pathways may bypass or overcome HIFs inhibition. Therefore, HIF-1 inhibitors may have to be combined with other targeted agents or conventional therapies to integrate hypoxia-targeting methods to get more reasonable results. Focusing research attention on these questions would thus not only be very beneficial for understanding the multifaceted roles of hypoxia on the hallmarks of human cancers but also facilitate the rational design of combination therapies to target hypoxia for cancer treatment.

ACKNOWLEDGMENTS

We apologize to those research groups whose work was not included in this review due to space limitation. We would like to acknowledge Shideng Bao (University of Colorado at Denver, Colorado), Li Yuan and reviewers for critical comments. This work was supported by the grants from the National Natural Science Foundation of China (No. 30400239, No. 30570935, No. 30871242), NCETXMU and a Berkeley Scholar Fellowship to G.O.

REFERENCES

Aragones J, Fraisl P, Baes M, Carmeliet P. 2009. Oxygen sensors at the crossroad of metabolism. *Cell Metab* 9:11–22.

Artandi SE, DePinho RA. 2000. A critical role for telomeres in suppressing and facilitating carcinogenesis. *Curr Opin Genet Dev* 10:39–46.

Bao S, Tibbetts RS, Brumbaugh KM, Fang Y, Richardson DA, Ali A, Chen SM, Abraham RT, Wang XF. 2001. ATR/ATM-mediated phosphorylation of human Rad17 is required for genotoxic stress responses. *Nature* 411:969–974.

Bao S, Ouyang G, Bai X, Huang Z, Ma C, Liu M, Shao R, Anderson RM, Rich JN, Wang XF. 2004. Periostin potently promotes metastatic growth of colon cancer by augmenting cell survival via the Akt/PKB pathway. *Cancer Cell* 5:329–339.

Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, Dewhirst MW, Bigner DD, Rich JN. 2006. Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature* 444:756–760.

Beavon IR. 1999. Regulation of E-cadherin: Does hypoxia initiate the metastatic cascade? *Mol Pathol* 52:179–188.

Bertout JA, Patel SA, Simon MC. 2008. The impact of O₂ availability on human cancer. *Nat Rev Cancer* 8:967–975.

Bindra RS, Schaffer PJ, Meng A, Woo J, Maseide K, Roth ME, Lizardi P, Hedley DW, Bristow RG, Glazer PM. 2005. Alterations in DNA repair gene expression under hypoxia: Elucidating the mechanisms of hypoxia-induced genetic instability. *Ann NY Acad Sci* 1059:184–195.

Bindra RS, Crosby ME, Glazer PM. 2007. Regulation of DNA repair in hypoxic cancer cells. *Cancer Metastasis Rev* 26:249–260.

Blagosklonny MV. 2004. Prospective strategies to enforce selectively cell death in cancer cells. *Oncogene* 23:2967–2975.

Brahimi-Horn MC, Chiche J, Pouyssegur J. 2007. Hypoxia signalling controls metabolic demand. *Curr Opin Cell Biol* 19:223–229.

Bristow RG, Hill RP. 2008. Hypoxia and metabolism. Hypoxia, DNA repair and genetic instability. *Nat Rev Cancer* 8:180–192.

Campisi J. 2005a. Senescent cells, tumor suppression, and organismal aging: Good citizens, bad neighbors. *Cell* 120:513–522.

Campisi J. 2005b. Suppressing cancer: The importance of being senescent. *Science* 309:886–887.

Campisi J, d'Adda di Fagagna F. 2007. Cellular senescence: When bad things happen to good cells. *Nat Rev Mol Cell Biol* 8:729–740.

Camps C, Buffa FM, Colella S, Moore J, Sotiriou C, Sheldon H, Harris AL, Gleadow JM, Ragoussis J. 2008. hsa-miR-210 is induced by hypoxia and is an independent prognostic factor in breast cancer. *Clin Cancer Res* 14:1340–1348.

Cao Y, Li CY, Moeller BJ, Yu D, Zhao Y, Dreher MR, Shan S, Dewhirst MW. 2005. Observation of incipient tumor angiogenesis that is independent of hypoxia and hypoxia inducible factor-1 activation. *Cancer Res* 65:5498–5505.

Chambers AF, Groom AC, MacDonald IC. 2002. Dissemination and growth of cancer cells in metastatic sites. *Nat Rev Cancer* 2:563–572.

Chan N, Milosevic M, Bristow RG. 2007. Tumor hypoxia, DNA repair and prostate cancer progression: New targets and new therapies. *Future Oncol* 3:329–341.

Chandel NS, Simon MC. 2008. Hypoxia-inducible factor: Roles in development, physiology, and disease. *Cell Death Differ* 15:619–620.

Chang Q, Qin R, Huang T, Gao J, Feng Y. 2006. Effect of antisense hypoxia-inducible factor 1alpha on progression, metastasis, and chemosensitivity of pancreatic cancer. *Pancreas* 32:297–305.

Chen EY, Mazure NM, Cooper JA, Giaccia AJ. 2001. Hypoxia activates a platelet-derived growth factor receptor/phosphatidylinositol 3-kinase/Akt pathway that results in glycogen synthase kinase-3 inactivation. *Cancer Res* 61:2429–2433.

Coquelle A, Toledo F, Stern S, Bieth A, Debatisse M. 1998. A new role for hypoxia in tumor progression: Induction of fragile site triggering genomic rearrangements and formation of complex DMs and HSRs. *Mol Cell* 2:259–265.

Croker AK, Allan AL. 2008. Cancer stem cells: Implications for the progression and treatment of metastatic disease. *J Cell Mol Med* 12:374–390.

- Dang CV, Kim JW, Gao P, Yustein J. 2008. The interplay between MYC and HIF in cancer. *Nat Rev Cancer* 8:51–56.
- DeBerardinis RJ, Lum JJ, Hatzivassiliou G, Thompson CB. 2008. The biology of cancer: Metabolic reprogramming fuels cell growth and proliferation. *Cell Metab* 7:11–20.
- Denko NC. 2008. Hypoxia, HIF1 and glucose metabolism in the solid tumour. *Nat Rev Cancer* 8:705–713.
- Dewhirst MW, Cao Y, Moeller B. 2008. Cycling hypoxia and free radicals regulate angiogenesis and radiotherapy response. *Nat Rev Cancer* 8:425–437.
- Dong Z, Venkatchalam MA, Wang J, Patel Y, Saikumar P, Semenza GL, Force T, Nishiyama J. 2001. Up-regulation of apoptosis inhibitory protein IAP-2 by hypoxia. Hif-1-independent mechanisms. *J Biol Chem* 276:18702–18709.
- Ellis LM, Hicklin DJ. 2008. VEGF-targeted therapy: Mechanisms of anti-tumour activity. *Nat Rev Cancer* 8:579–591.
- Erler JT, Bennewith KL, Nicolau M, Dornhofer N, Kong C, Le QT, Chi JT, Jeffrey SS, Giaccia AJ. 2006. Lysyl oxidase is essential for hypoxia-induced metastasis. *Nature* 440:1222–1226.
- Fang JS, Gillies RD, Gatenby RA. 2008. Adaptation to hypoxia and acidosis in carcinogenesis and tumor progression. *Semin Cancer Biol* 18:330–337.
- Furuta E, Pai SK, Zhan R, Bandyopadhyay S, Watabe M, Mo YY, Hirota S, Hosobe S, Tsukada T, Miura K, Kamada S, Saito K, Iizumi M, Liu W, Ericsson J, Watabe K. 2008. Fatty acid synthase gene is up-regulated by hypoxia via activation of Akt and sterol regulatory element binding protein-1. *Cancer Res* 68:1003–1011.
- Gatenby RA, Gillies RJ. 2008. A microenvironmental model of carcinogenesis. *Nat Rev Cancer* 8:56–61.
- Gatenby RA, Smallbone K, Maini PK, Rose F, Averill J, Nagle RB, Worrall L, Gillies RJ. 2007. Cellular adaptations to hypoxia and acidosis during somatic evolution of breast cancer. *Br J Cancer* 97:646–653.
- Gibson SL, Bindra RS, Glazer PM. 2005. Hypoxia-induced phosphorylation of Chk2 in an ataxia telangiectasia mutated-dependent manner. *Cancer Res* 65:10734–10741.
- Gibson SL, Bindra RS, Glazer PM. 2006. CHK2-dependent phosphorylation of BRCA1 in hypoxia. *Radiat Res* 166:646–651.
- Gillies RJ, Gatenby RA. 2007. Hypoxia and adaptive landscapes in the evolution of carcinogenesis. *Cancer Metastasis Rev* 26:311–317.
- Graeber TG, Osmanian C, Jacks T, Housman DE, Koch CJ, Lowe SW, Giaccia AJ. 1996. Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumours. *Nature* 379:88–91.
- Hahn WC, Weinberg RA. 2002. Modelling the molecular circuitry of cancer. *Nat Rev Cancer* 2:331–341.
- Hammond EM, Denko NC, Dorie MJ, Abraham RT, Giaccia AJ. 2002. Hypoxia links ATR and p53 through replication arrest. *Mol Cell Biol* 22:1834–1843.
- Hammond EM, Dorie MJ, Giaccia AJ. 2003. ATR/ATM targets are phosphorylated by ATR in response to hypoxia and ATM in response to reoxygenation. *J Biol Chem* 278:12207–12213.
- Hanahan D, Weinberg RA. 2000. The hallmarks of cancer. *Cell* 100:57–70.
- Harris AL. 2002. Hypoxia—A key regulatory factor in tumour growth. *Nat Rev Cancer* 2:38–47.
- Hockel M, Vaupel P. 2001. Tumor hypoxia: Definitions and current clinical, biologic, and molecular aspects. *J Natl Cancer Inst* 93:266–276.
- Holash J, Maisonpierre PC, Compton D, Boland P, Alexander CR, Zagzag D, Yancopoulos GD, Wiegand SJ. 1999. Vessel cooption, regression, and growth in tumors mediated by angiopoietins and VEGF. *Science* 284:1994–1998.
- Huang LE, Bindra RS, Glazer PM, Harris AL. 2007. Hypoxia-induced genetic instability—A calculated mechanism underlying tumor progression. *J Mol Med* 85:139–148.
- Imai T, Horiuchi A, Wang C, Oka K, Ohira S, Nikaido T, Konishi I. 2003. Hypoxia attenuates the expression of E-cadherin via up-regulation of SNAI1 in ovarian carcinoma cells. *Am J Pathol* 163:1437–1447.
- Ivan M, Harris AL, Martelli F, Kulshreshtha R. 2008. Hypoxia response and microRNAs: No longer two separate worlds. *J Cell Mol Med* 12:1426–1431.
- Jaakkola P, Mole DR, Tian YM, Wilson MI, Gielbert J, Gaskell SJ, Kriegsheim A, Hebestreit HF, Mukherji M, Schofield CJ, Maxwell PH, Pugh CW, Ratcliffe PJ. 2001. Targeting of HIF- α to the von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science* 292:468–472.
- Jiang YG, Luo Y, He DL, Li X, Zhang LL, Peng T, Li MC, Lin YH. 2007. Role of Wnt/ β -catenin signaling pathway in epithelial-mesenchymal transition of human prostate cancer induced by hypoxia-inducible factor-1 α . *Int J Urol* 14:1034–1039.
- Joyce JA, Pollard JW. 2009. Microenvironmental regulation of metastasis. *Nat Rev Cancer* 9:239–252.
- Kaelin WG, Jr., Ratcliffe PJ. 2008. Oxygen sensing by metazoans: The central role of the HIF hydroxylase pathway. *Mol Cell* 30:393–402.
- Kaidi A, Williams AC, Paraskeva C. 2007. Interaction between β -catenin and HIF-1 promotes cellular adaptation to hypoxia. *Nat Cell Biol* 9:210–217.
- Kaluz S, Kaluzova M, Stanbridge EJ. 2008. Does inhibition of degradation of hypoxia-inducible factor (HIF) α always lead to activation of HIF? Lessons learnt from the effect of proteasomal inhibition on HIF activity. *J Cell Biochem* 104:536–544.
- Kato H, Inoue T, Asanoma K, Nishimura C, Matsuda T, Wake N. 2006. Induction of human endometrial cancer cell senescence through modulation of HIF-1 α activity by EGLN1. *Int J Cancer* 118:1144–1153.
- Keith B, Simon MC. 2007. Hypoxia-inducible factors, stem cells, and cancer. *Cell* 129:465–472.
- Kim WY, Sharpless NE. 2008. VHL inactivation: A new road to senescence. *Cancer Cell* 13:295–297.
- Kim JW, Tchernyshyov I, Semenza GL, Dang CV. 2006. HIF-1-mediated expression of pyruvate dehydrogenase kinase: A metabolic switch required for cellular adaptation to hypoxia. *Cell Metab* 3:177–185.
- Kim JW, Gao P, Dang CV. 2007. Effects of hypoxia on tumor metabolism. *Cancer Metastasis Rev* 26:291–298.
- Kondo A, Safaei R, Mishima M, Niedner H, Lin X, Howell SB. 2001. Hypoxia-induced enrichment and mutagenesis of cells that have lost DNA mismatch repair. *Cancer Res* 61:7603–7607.
- Koshiji M, To KK, Hammer S, Kumamoto K, Harris AL, Modrich P, Huang LE. 2005. HIF-1 α induces genetic instability by transcriptionally down-regulating MutS α expression. *Mol Cell* 17:793–803.
- Krishnamachary B, Berg-Dixon S, Kelly B, Agani F, Feldser D, Ferreira G, Iyer N, LaRusch J, Pak B, Taghavi P, Semenza GL. 2003. Regulation of colon carcinoma cell invasion by hypoxia-inducible factor 1. *Cancer Res* 63:1138–1143.
- Krishnamachary B, Zagzag D, Nagasawa H, Rainey K, Okuyama H, Baek JH, Semenza GL. 2006. Hypoxia-inducible factor-1-dependent repression of E-cadherin in von Hippel-Lindau tumor suppressor-null renal cell carcinoma mediated by TCF3, ZFH1A, and ZFH1B. *Cancer Res* 66:2725–2731.
- Kroemer G, Pouyssegur J. 2008. Tumor cell metabolism: Cancer's Achilles' heel. *Cancer Cell* 13:472–482.
- Kulshreshtha R, Ferracin M, Wojcik SE, Garzon R, Alder H, Agosto-Perez FJ, Davuluri R, Liu CG, Croce CM, Negrini M, Calin GA, Ivan M. 2007. A microRNA signature of hypoxia. *Mol Cell Biol* 27:1859–1867.
- Kunz M, Ibrahim S, Koczan D, Thiesen HJ, Kohler HJ, Acker T, Plate KH, Ludwig S, Rapp UR, Bocker EB, van Muijen GN, Flory E, Gross G. 2001. Activation of c-Jun NH₂-terminal kinase/stress-activated protein kinase (JNK/SAPK) is critical for hypoxia-induced apoptosis of human malignant melanoma. *Cell Growth Differ* 12:137–145.
- Laderoute KR, Alarcon RM, Brody MD, Calaoagan JM, Chen EY, Knapp AM, Yun Z, Denko NC, Giaccia AJ. 2000. Opposing effects of hypoxia on

- expression of the angiogenic inhibitor thrombospondin 1 and the angiogenic inducer vascular endothelial growth factor. *Clin Cancer Res* 6:2941–2950.
- Lando D, Peet DJ, Gorman JJ, Whelan DA, Whitelaw ML, Bruick RK. 2002. FIH-1 is an asparaginyl hydroxylase enzyme that regulates the transcriptional activity of hypoxia-inducible factor. *Genes Dev* 16:1466–1471.
- Le QT, Courter D. 2008. Clinical biomarkers for hypoxia targeting. *Cancer Metastasis Rev* 27:351–362.
- Lewis C, Murdoch C. 2005. Macrophage responses to hypoxia: Implications for tumor progression and anti-cancer therapies. *Am J Pathol* 167:627–635.
- Liao D, Corle C, Seagroves TN, Johnson RS. 2007. Hypoxia-inducible factor-1alpha is a key regulator of metastasis in a transgenic model of cancer initiation and progression. *Cancer Res* 67:563–572.
- Lisy K, Peet DJ. 2008. Turn me on: Regulating HIF transcriptional activity. *Cell Death Differ* 15:642–649.
- Lopez-Maury L, Marguerat S, Bahler J. 2008. Tuning gene expression to changing environments: From rapid responses to evolutionary adaptation. *Nat Rev Genet* 9:583–593.
- Lukashev D, Ohta A, Sitkovsky M. 2007. Hypoxia-dependent anti-inflammatory pathways in protection of cancerous tissues. *Cancer Metastasis Rev* 26:273–279.
- Lundgren K, Holm C, Landberg G. 2007. Hypoxia and breast cancer: Prognostic and therapeutic implications. *Cell Mol Life Sci* 64:3233–3247.
- Ma C, Rong Y, Radloff DR, Datto MB, Centeno B, Bao S, Cheng AW, Lin F, Jiang S, Yeatman TJ, Wang XF. 2008. Extracellular matrix protein betaig-h3/TGFBI promotes metastasis of colon cancer by enhancing cell extravasation. *Genes Dev* 22:308–321.
- Maisonpierre PC, Suri C, Jones PF, Bartunkova S, Wiegand SJ, Radziejewski C, Compton D, McClain J, Aldrich TH, Papadopoulos N, Daly TJ, Davis S, Sato TN, Yancopoulos GD. 1997. Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis. *Science* 277:55–60.
- Makino Y, Nakamura H, Ikeda E, Ohnuma K, Yamauchi K, Yabe Y, Poellinger L, Okada Y, Morimoto C, Tanaka H. 2003. Hypoxia-inducible factor regulates survival of antigen receptor-driven T cells. *J Immunol* 171:6534–6540.
- Maxwell PH, Pugh CW, Ratcliffe PJ. 2001. Activation of the HIF pathway in cancer. *Curr Opin Genet Dev* 11:293–299.
- Murata Y, Ohteki T, Koyasu S, Hamuro J. 2002. IFN-gamma and pro-inflammatory cytokine production by antigen-presenting cells is dictated by intracellular thiol redox status regulated by oxygen tension. *Eur J Immunol* 32:2866–2873.
- Nelson DA, Tan TT, Rabson AB, Anderson D, Degenhardt K, White E. 2004. Hypoxia and defective apoptosis drive genomic instability and tumorigenesis. *Genes Dev* 18:2095–2107.
- Nguyen DX, Bos PD, Massague J. 2009. Metastasis: From dissemination to organ-specific colonization. *Nat Rev Cancer* 9:274–284.
- Ohh M, Park CW, Ivan M, Hoffman MA, Kim TY, Huang LE, Pavletich N, Chau V, Kaelin WG. 2000. Ubiquitination of hypoxia-inducible factor requires direct binding to the beta-domain of the von Hippel-Lindau protein. *Nat Cell Biol* 2:423–427.
- Ouyang G, Liu M, Ruan K, Song G, Mao Y, Bao S. 2009. Upregulated expression of periostin by hypoxia in non-small-cell lung cancer cells promotes cell survival via the Akt/PKB pathway. *Cancer Lett* [Epub ahead of print].
- Papandreou I, Cairns RA, Fontana L, Lim AL, Denko NC. 2006. HIF-1 mediates adaptation to hypoxia by actively downregulating mitochondrial oxygen consumption. *Cell Metab* 3:187–197.
- Peinado H, Olmeda D, Cano A. 2007. Snail, Zeb and bHLH factors in tumour progression: An alliance against the epithelial phenotype? *Nat Rev Cancer* 7:415–428.
- Polyak K, Weinberg RA. 2009. Transitions between epithelial and mesenchymal states: Acquisition of malignant and stem cell traits. *Nat Rev Cancer* 9:265–273.
- Pouyssegur J, Dayan F, Mazure NM. 2006. Hypoxia signalling in cancer and approaches to enforce tumour regression. *Nature* 441:437–443.
- Powis G, Kirkpatrick L. 2004. Hypoxia inducible factor-1alpha as a cancer drug target. *Mol Cancer Ther* 3:647–654.
- Rangaswami H, Bulbule A, Kundu GC. 2006. Osteopontin: Role in cell signaling and cancer progression. *Trends Cell Biol* 16:79–87.
- Rich JN, Bao S. 2007. Chemotherapy and cancer stem cells. *Cell Stem Cell* 1:353–355.
- Richard DE, Berra E, Gothie E, Roux D, Pouyssegur J. 1999. p42/p44 mitogen-activated protein kinases phosphorylate hypoxia-inducible factor 1alpha (HIF-1alpha) and enhance the transcriptional activity of HIF-1. *J Biol Chem* 274:32631–32637.
- Rius J, Guma M, Schachtrup C, Akassoglou K, Zinkernagel AS, Nizet V, Johnson RS, Haddad GG, Karin M. 2008. NF-kappaB links innate immunity to the hypoxic response through transcriptional regulation of HIF-1alpha. *Nature* 453:807–811.
- Roskelley CD, Bissell MJ. 2002. The dominance of the microenvironment in breast and ovarian cancer. *Semin Cancer Biol* 12:97–104.
- Ruan K, Bao S, Ouyang G. 2009. The multifaceted role of periostin in tumorigenesis. *Cell Mol Life Sci* [Epub ahead of print].
- Sahlgren C, Gustafsson MV, Jin S, Poellinger L, Lendahl U. 2008. Notch signaling mediates hypoxia-induced tumor cell migration and invasion. *Proc Natl Acad Sci USA* 105:6392–6397.
- Schmid T, Zhou J, Brune B. 2004. HIF-1 and p53: Communication of transcription factors under hypoxia. *J Cell Mol Med* 8:423–431.
- Seimiya H, Tanji M, Oh-hara T, Tomida A, Naasani I, Tsuruo T. 1999. Hypoxia up-regulates telomerase activity via mitogen-activated protein kinase signaling in human solid tumor cells. *Biochem Biophys Res Commun* 260:365–370.
- Semenza GL. 2002. HIF-1 and tumor progression: Pathophysiology and therapeutics. *Trends Mol Med* 8:S62–S67.
- Semenza GL. 2003. Targeting HIF-1 for cancer therapy. *Nat Rev Cancer* 3:721–732.
- Semenza GL. 2007a. Evaluation of HIF-1 inhibitors as anticancer agents. *Drug Discov Today* 12:853–859.
- Semenza GL. 2007b. Life with oxygen. *Science* 318:62–64.
- Shen C, Zhou Y, Zhan J, Reske SN, Buck AK. 2008. Chromosome instability and tumor lethality suppression in carcinogenesis. *J Cell Biochem* 105:1327–1341.
- Shi Q, Bao S, Maxwell JA, Reese ED, Friedman HS, Bigner DD, Wang XF, Rich JN. 2004. Secreted protein acidic, rich in cysteine (SPARC), mediates cellular survival of gliomas through AKT activation. *J Biol Chem* 279:52200–52209.
- Simon MC, Keith B. 2008. The role of oxygen availability in embryonic development and stem cell function. *Nat Rev Mol Cell Biol* 9:285–296.
- Sitkovsky M, Lukashev D. 2005. Regulation of immune cells by local-tissue oxygen tension: HIF1 alpha and adenosine receptors. *Nat Rev Immunol* 5:712–721.
- Soengas MS, Alarcon RM, Yoshida H, Giaccia AJ, Hakem R, Mak TW, Lowe SW. 1999. Apaf-1 and caspase-9 in p53-dependent apoptosis and tumor inhibition. *Science* 284:156–159.
- Song G, Ouyang G, Bao S. 2005. The activation of Akt/PKB signaling pathway and cell survival. *J Cell Mol Med* 9:59–71.
- Song G, Cai QF, Mao YB, Ming YL, Bao SD, Ouyang GL. 2008a. Osteopontin promotes ovarian cancer progression and cell survival and increases HIF-1alpha expression through the PI3-K/Akt pathway. *Cancer Sci* 99:1901–1907.
- Song G, Ming Y, Mao Y, Bao S, Ouyang G. 2008b. Osteopontin Prevents Curcumin-Induced Apoptosis and Promotes Survival Through Akt Activation via $\alpha\beta_3$ Integrins in Human Gastric Cancer Cells. *Exp Biol Med* (Maywood) 233:1537–1545.

- Sowter HM, Ratcliffe PJ, Watson P, Greenberg AH, Harris AL. 2001. HIF-1-dependent regulation of hypoxic induction of the cell death factors BNIP3 and NIX in human tumors. *Cancer Res* 61:6669–6673.
- Suzuki H, Tomida A, Tsuruo T. 2001. Dephosphorylated hypoxia-inducible factor 1alpha as a mediator of p53-dependent apoptosis during hypoxia. *Oncogene* 20:5779–5788.
- Talks KL, Turley H, Gatter KC, Maxwell PH, Pugh CW, Ratcliffe PJ, Harris AL. 2000. The expression and distribution of the hypoxia-inducible factors HIF-1alpha and HIF-2alpha in normal human tissues, cancers, and tumor-associated macrophages. *Am J Pathol* 157:411–421.
- Wai PY, Kuo PC. 2008. Osteopontin: Regulation in tumor metastasis. *Cancer Metastasis Rev* 27:103–118.
- Walsh S, Gill C, O'Neill A, Fitzpatrick JM, Watson RW. 2009. Hypoxia increases normal prostate epithelial cell resistance to receptor-mediated apoptosis via AKT activation. *Int J Cancer* 124:1871–1878.
- Winner M, Leng L, Zundel W, Mitchell RA. 2007. Macrophage migration inhibitory factor manipulation and evaluation in tumoral hypoxic adaptation. *Methods Enzymol* 435:355–369.
- Wouters BG, Koritzinsky M. 2008. Hypoxia signalling through mTOR and the unfolded protein response in cancer. *Nat Rev Cancer* 8:851–864.
- Yang J, Mani SA, Donaher JL, Ramaswamy S, Itzykson RA, Come C, Savagner P, Gitelman I, Richardson A, Weinberg RA. 2004. Twist, a master regulator of morphogenesis, plays an essential role in tumor metastasis. *Cell* 117:927–939.
- Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, Teng SC, Wu KJ. 2008. Direct regulation of TWIST by HIF-1alpha promotes metastasis. *Nat Cell Biol* 10:295–305.
- Yaswen P, Campisi J. 2007. Oncogene-induced senescence pathways weave an intricate tapestry. *Cell* 128:233–234.
- Young AP, Schlisio S, Minamishima YA, Zhang Q, Li L, Grisanzio C, Signoretti S, Kaelin WG, Jr. 2008. VHL loss activates a HIF-independent senescence programme mediated by Rb and p400. *Nat Cell Biol* 10:361–369.
- Zeng R, Yao Y, Han M, Zhao X, Liu XC, Wei J, Luo Y, Zhang J, Zhou J, Wang S, Ma D, Xu G. 2008. Biliverdin reductase mediates hypoxia-induced EMT via PI3-kinase and Akt. *J Am Soc Nephrol* 19:380–387.
- Zhang L, Hill RP. 2004. Hypoxia enhances metastatic efficiency by up-regulating Mdm2 in KHT cells and increasing resistance to apoptosis. *Cancer Res* 64:4180–4189.
- Zhang H, Gao P, Fukuda R, Kumar G, Krishnamachary B, Zeller KI, Dang CV, Semenza GL. 2007. HIF-1 inhibits mitochondrial biogenesis and cellular respiration in VHL-deficient renal cell carcinoma by repression of C-MYC activity. *Cancer Cell* 11:407–420.
- Zhong H, De Marzo AM, Laughner E, Lim M, Hilton DA, Zagzag D, Buechler P, Isaacs WB, Semenza GL, Simons JW. 1999. Overexpression of hypoxia-inducible factor 1alpha in common human cancers and their metastases. *Cancer Res* 59:5830–5835.
- Zundel W, Schindler C, Haas-Kogan D, Koong A, Kaper F, Chen E, Gottschalk AR, Ryan HE, Johnson RS, Jefferson AB, Stokoe D, Giaccia AJ. 2000. Loss of PTEN facilitates HIF-1-mediated gene expression. *Genes Dev* 14:391–396.